



Administration of CD34⁺ cells after stroke enhances neurogenesis via angiogenesis in a mouse model

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Thrombo-occlusive cerebrovascular disease resulting in stroke and permanent neuronal loss is an important cause of morbidity and mortality. Because of the unique properties of cerebral vasculature and the limited reparative capability of neuronal tissue, it has been difficult to devise effective neuroprotective therapies in cerebral ischemia. Our results demonstrate that systemic administration of human cord blood–derived CD34⁺ cells to immunocompromised mice subjected to stroke 48 hours earlier induces neovascularization in the ischemic zone and provides a favorable environment for neuronal regeneration. Endogenous neurogenesis, suppressed by an antiangiogenic agent, is accelerated as a result of enhanced migration of neuronal progenitor cells to the damaged area, followed by their maturation and functional recovery. Our data suggest an essential role for CD34⁺ cells in promoting directly or indirectly an environment conducive to neovascularization of ischemic brain so that neuronal regeneration can proceed.

Introduction

Thrombo-occlusive atherosclerotic cardiovascular disease is a major cause of death and disability in developed countries. In the acute phase, therapeutic maneuvers include fibrinolytic therapy to restore blood flow to the ischemic site. In the longer term, formation of new blood vessels is necessary to fully supply tissue metabolic and functional requirements. Although it had been assumed that postnatal development of neovessels resulted only from outgrowth of pre-existing vasculature, it has become evident that circulating endothelial progenitor cells (EPCs), contained in a CD34⁺ cell population enriched in cord blood, have the capacity to participate in neovascularization of ischemic tissues (1, 2). Thus, a new strategy proposed for enhancing recovery due to ischemic stress is administration of EPCs to stimulate formation of neovasculature. In this context, recent reports have demonstrated that infusion of EPCs results in their incorporation into neovasculature at the ischemic site and limitation of tissue damage in animal models (3). Furthermore, human CD34⁺ cells were shown to secrete numerous angiogenic factors, including VEGF, HGF, and IGF-1 (4). On the basis of these observations, clinical trials of cell transplantation in hindlimb (5, 6) and cardiac ischemia (7) have been initiated with promising results.

Nonstandard abbreviations used: anterior cerebral artery (ACA); cerebral blood flow (CBF); chloromethylbenzamide (CM-Dil); doublecortin (DCX); endothelial progenitor cell (EPC); erythropoietin (EPO); fetal liver kinase-1 (Flk-1); high-power field (HPF); middle cerebral artery (MCA); neuronal progenitor cell (NPC); neuron-specific nuclear protein (NeuN); phycoerythrin (PE); polysialylated neuronal cell adhesion molecule (PSA-NCAM); subventricular zone (SVZ); 2,3,5-triphenyltetrazolium (TTC).

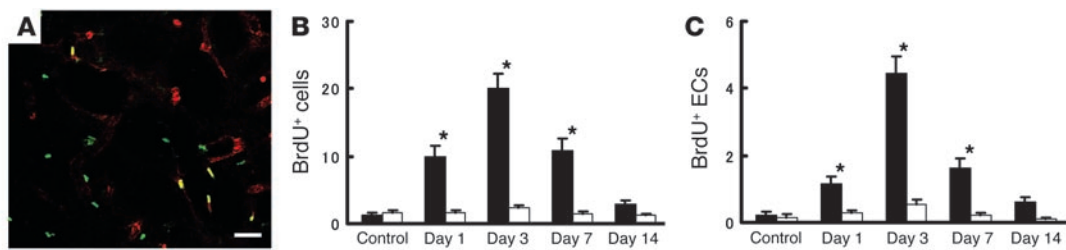
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Stroke is another setting of occlusive thromboatherosclerotic disease in which acceleration of angiogenesis might be expected to enhance the outcome. Despite the improvement of poststroke neurological outcome by administration of human cord blood cells (8) or bone marrow–derived cells (9) (both potentially a rich sources of stem cells including CD34⁺ cells) in rodent models, few of the administered cells could be demonstrated in brain parenchyma expressing neuronal markers, raising a question as to the underlying mechanism. The results of our study demonstrate that systemic administration of human CD34⁺ cells to immunocompromised mice subjected to stroke 48 hours earlier accelerates neovascularization of the ischemic zone. Such a rich vascular environment, along with generation of other nurturing neuronal mediators by CD34⁺ cells, such as VEGF, FGF2, and IGF-1 (10–12), enhances subsequent neuronal regeneration; endogenous neurogenesis is accelerated as neuronal progenitors migrate to the damaged area, followed by their maturation and survival when CD34⁺ cells have stimulated the formation of increased vascular channels. In contrast, in the presence of an antiangiogenic agent, the beneficial effect of CD34⁺ cells was lost. Our results provide the first direct link between vasculogenesis and neurogenesis in the repair of ischemic brain lesions.

Results

Induction of stroke and proliferation of endothelial cells in situ. A reproducible model of stroke in the middle cerebral artery (MCA) cortex, sparing the striatum, was developed in SCID mice by permanent ligation of the M1 distal portion of the left MCA. Subsequent infusion of carbon black showed strongly decreased staining in the affected area. Nonviability of affected tissue was confirmed by 2,3,5-triphenyltetrazolium (TTC) staining. Values of cortical width index (see Methods section) were highly reproducible (–0.34–0.36)

**Figure 1**

Endothelial proliferation in situ after stroke. On days 1, 3, 7, and 14 after stroke, the number of proliferating cells (BrdU⁺) and proliferating endothelial cells (co-staining for BrdU and CD31) was determined in the left cortical area of 1–1.5 mm distal from the midline. (A) Immunohistological analysis of proliferating cells labeled with BrdU (green), anti-mouse CD31 IgG (red), and both (yellow). The number of cells visualized with BrdU (B) and the subpopulation BrdU⁺ cells also displaying mouse CD31 (i.e., double positives) (C) are shown. Ten HPFs were evaluated for each animal ($n = 6$ per group) by two investigators blinded to the experimental protocol. Note in C, cells displaying mouse CD31 are termed endothelial cells (ECs). Black bars, ipsilateral; white bars, contralateral. * $P < 0.05$ versus control. Scale bar: 30 μm .

over the 12-week experimental period. Survival in this stroke model was greater than 95%, and no seizures were observed.

To estimate the optimal time to administer human CD34⁺ cells, proliferation of endothelial cells in vasculature of the penumbral region (at the leading edge of viable tissue) was assessed by in vivo BrdU labeling. Sections were visualized with antibody to BrdU and mouse-specific antibody to CD31 by confocal microscopy. Cellular profiles co-staining for both markers were considered proliferating endothelial cells (Figure 1A). On days 1 and 3 after stroke, a subpopulation of BrdU⁺ cells also stained with mouse CD31, indicating an endothelial origin of this signal (Figure 1, B and C). By day 7, although endothelial proliferation continued, it had begun to decrease. In contrast, BrdU-labeled cells were present in a constant, small amount on the contralateral (nonstroke) side (Figure 1, B and C). These data indicated that administration of CD34⁺ cells on day 2 after stroke would buttress the endogenous proliferative component of the vascular response to cerebral ischemia.

Administration of CD34⁺ cells after stroke. Human CD34⁺ cells (95% pure CD34⁺ cells) isolated from human cord blood or control cells (CD34⁻ cells with <0.2% CD34⁺ cells, also from human cord blood) were administered intravenously via tail vein 48 hours after stroke. Analysis of cell surface markers revealed that $1.5 \pm 0.1\%$ and $0.9 \pm 0.1\%$ of the CD34⁺ cell population expressed the endothelial lineage markers fetal liver kinase-1 (Flk-1) (1) and P1H12 (13), respectively ($n = 4$). The effect of CD34⁺ cells was evident within 24 hours of their transplantation. Labeling vasculature by infusion of carbon black ink demonstrated neovasculature at the border of the MCA and anterior cerebral artery (ACA) cortex (staining with TTC demarcates viable and nonviable tissue) in animals treated with CD34⁺ cells (Figure 2, A and B), compared with those receiving CD34⁻ cells (Figure 2C) or PBS alone (Figure 2D). Determination of the angiographic score confirmed the impression of increased neovasculature in animals transplanted with CD34⁺ cells, compared with other groups (Figure 2E). To evaluate vascular activation in affected cerebral vessels, we used mouse-specific antibody to CD13, an antigen expressed by endothelial cells in angiogenic, but not quiescent, vasculature (14). Visualization of mouse CD13 in brain sections 24 hours after cell transplantation showed that cells bearing this activated endothelial marker were most evident in sections from mice treated with CD34⁺ (Figure 2F), compared with those receiving CD34⁻ cells (Figure 2G) or PBS (Figure 2H). Increased density of vasculature in the ischemic territory of animals treated with CD34⁺ cells translated to significantly enhanced cerebral blood flow (CBF) (Figure 2I).

To analyze the effect of subpopulations within the general CD34⁺ cell population, we compared the effect of poststroke transplantation of the same number of CD34⁺ cells (containing Flk-1⁻ and Flk-1⁺ cells) with CD34⁺/Flk-1⁻ cells on vascular activation and neovasculature. Brain tissue was examined 7 days after cell transplantation, because EPCs are known to incorporate into capillary walls at ischemic sites by this time point after the ischemic episode (2). FACS analysis confirmed that the CD34⁺/Flk-1⁻ population contained less than 0.1% Flk-1⁺ cells ($n = 4$). On the basis of CD13 staining (using the same mouse-specific antibody mentioned earlier), there was similar activation of endogenous endothelium after transplantation of either CD34⁺ cells (including both Flk-1⁻ and Flk-1⁺ subpopulations) and CD34⁺/Flk-1⁻ cells (not shown). Although neovasculature was observed at the border of the MCA and ACA cortex in animals treated with CD34⁺ cells (Figure 2J) and CD34⁺/Flk-1⁻ cells (Figure 2K), mice treated with CD34⁺ cells displayed increased neovasculature based on angiographic score (scores of 22 ± 3 and 13 ± 2 , for CD34⁺ and CD34⁺/Flk-1⁻ cells, respectively; $P < 0.05$, $n = 6$).

Transplantation of CD34⁺ cells and poststroke functional recovery. Stroke causes motor deficits and behavioral abnormalities (15). Dysfunction of the cortex is closely linked to disinhibition of behavior (16). Compared with sham-operated controls, mice that received CD34⁻ cells or PBS displayed significant behavioral abnormalities on day 90 after cell transplantation ($n = 12$, for each group). Rearing counts under lighted conditions were 8.4 ± 0.8 (PBS), 8.7 ± 0.5 (CD34⁻ cells), 4.2 ± 0.5 (CD34⁺ cells), and 3.0 ± 0.5 (sham-operated controls) for each of the groups. Counts of locomotion were 5.1 ± 0.4 (PBS), 5.0 ± 0.4 (CD34⁻ cells), 3.7 ± 0.4 (CD34⁺ cells), and 3.3 ± 0.4 (sham). ANOVA revealed hyperactivity with respect to both rearing ($P < 0.01$) and locomotion ($P < 0.01$) in the CD34⁻ cell- and PBS-treated groups, compared with sham-operated controls. In contrast, mice treated with CD34⁺ cells showed no significant hyperactivity compared with sham-operated controls ($P > 0.05$), and displayed significant improvement in both behavioral tests compared with animals that received PBS or CD34⁻ cells ($P < 0.05$). Mice treated with CD34⁻ cells or PBS after stroke showed loss of this “dark” response, with respect to rearing ($P > 0.05$) and locomotion ($P > 0.05$). In contrast, animals treated with CD34⁺ cells displayed the expected increase in rearing and locomotion in the darkness ($P < 0.01$).

Using another behavioral paradigm, excessive startle consequent to auditory stimulation was observed in poststroke animals treated with CD34⁻ cells and PBS. Startle amplitudes were 0.9 ± 0.1 volts (PBS), 0.8 ± 0.1 (CD34⁻ cells), 0.5 ± 0.1 (CD34⁺

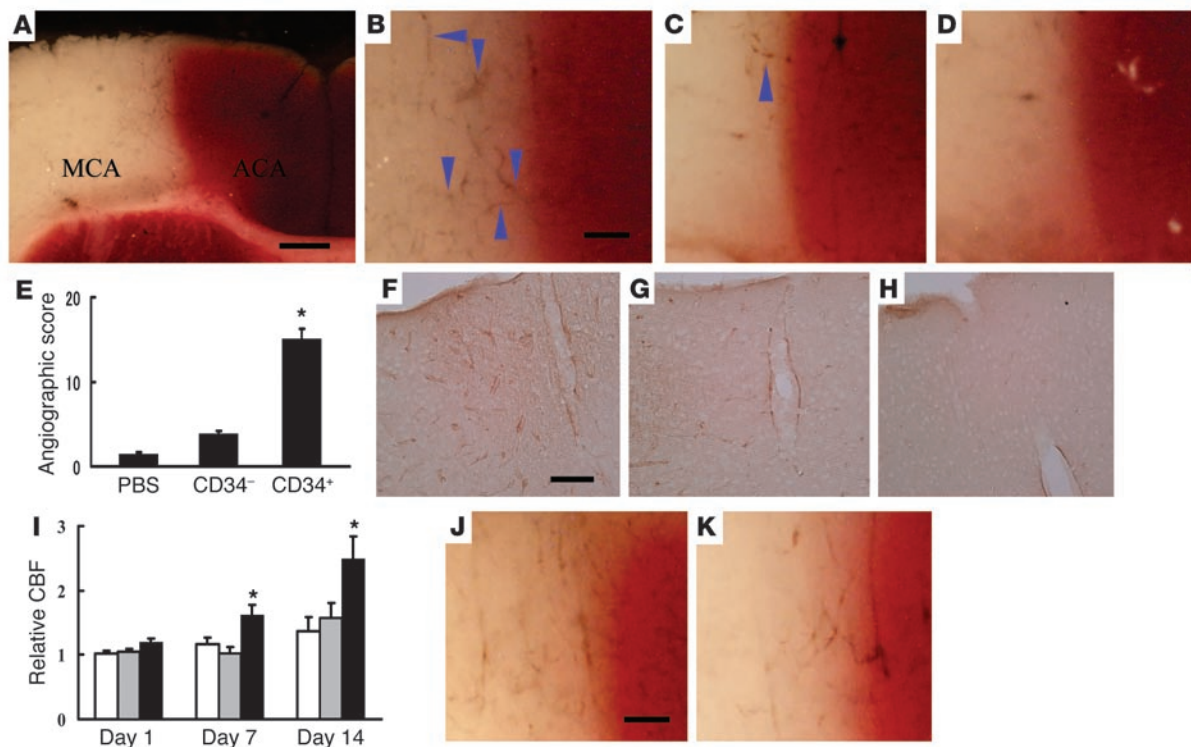


Figure 2 Transplantation of CD34⁺ cells after stroke accelerates neovascularization. (A–D) Mice subjected to stroke received CD34⁺ cells (A and B), CD34⁻ cells (C), or PBS alone (D) on day 2. Animals were infused with carbon black ink and killed at 24 hours after cell transplantations. Sections were stained with TTC. Neovascularization was noted at the border zone between the ACA and MCA areas (arrowheads show microvessels), especially in animals treated with CD34⁺ cells compared with those receiving CD34⁻ cells or PBS alone. (E) An angiographic score for each experimental condition based on analysis of 6 mice per group. (F–H) Activated endothelial cells were observed with antibody specific for mouse CD13 in the ACA area. F: CD34⁺ cells; G: CD34⁻ cells; H: PBS. (I) CBF was measured in the MCA area just outside of the penumbra, and values in animals treated with CD34⁺ cells (black bars), CD34⁻ cells (gray bars), or PBS (white bars) were compared with values before cell transplantation at times corresponding to days 1, 7, and 14 after cell transplantation (*n* = 6 per group). Data shown are relative CBF versus day the measurement was performed. (J and K) Labeling vasculature by infusion of carbon black ink demonstrated neovasculture at the border of the MCA and ACA cortex in animals treated with CD34⁺ cells (J) and CD34⁺/Flk-1⁻ cells (K) on day 7 after cell transplantation. Scale bars: 0.5 mm (A) and 0.1 mm (B, F, and J). **P* < 0.05 versus PBS.

cells), and 0.4 ± 0.1 (sham) on day 90. ANOVA revealed hyperactivity in the CD34⁻ cell- and PBS-treated groups, compared with sham-operated controls (*P* < 0.05). In contrast, mice treated with CD34⁺ cells showed no significant hyperactivity compared with sham-operated controls (*P* > 0.05). Shortening of the latency period with fear conditioning was also observed in poststroke animals treated with CD34⁻ cells and PBS compared with sham-operated controls (*P* < 0.05). In contrast, mice treated with CD34⁺ cells showed no significant hyperactivity (*P* > 0.05). Similar behavioral abnormalities were also observed on day 35 after cell transplantation. Since ischemic injury in our stroke model was limited to the cortex (hippocampus and striatum were intact), it was not surprising that spatial learning (water maze testing) and passive avoidance remained unchanged and comparable in all groups when assessed 5 weeks after stroke (data not shown). Mice showed rapid recovery from focal motor deficits, and by day 16 after stroke no motor deficits were detected using a modified three-point scale (17) (data not shown).

Effect of poststroke CD34⁺ cell transplantation on endogenous neurogenesis and cortical expansion. These results suggested that cortical function and neoangiogenesis, potentially supporting integrity of the MCA cortex, might be better maintained in animals treated with CD34⁺

cells after stroke. Such maintenance of cortical integrity might reflect CD34⁺ cell-mediated enhancement of the survival of ischemic neurons or acceleration of endogenous neurogenesis. Representative brains from poststroke animals in each group (treated with PBS alone, CD34⁻ cells, or CD34⁺ cells) are shown in Figure 3, A–F. There was a prominent increase in cortical width index in animals receiving CD34⁺ cells at 14, 35, and 90 days after cell transplantation (Figure 3G). Thus, cortical expansion in animals treated with CD34⁺ cells was most likely to reflect the result of ongoing neurogenesis, rather than maintenance of the integrity of ischemic neurons from the time of induction of stroke.

Cell trace analysis with a fluorescent dye revealed that labeled transplanted CD34⁺ cells were not co-stained with antibody to a neuronal (neuron-specific nuclear protein; NeuN) or an astrocyte (glial fibrillary acidic protein) marker (not shown). However, activation of endogenous neurogenesis was observed after stroke, and this was accentuated in mice receiving CD34⁺ cells. On day 7 after administration of CD34⁺ cells, cells expressing the polysialylated neuronal cell adhesion molecule (PSA-NCAM), a marker of migrating neuronal progenitor cells (NPCs) (18), were observed in the subventricular zone (SVZ), and migration of these NPCs toward the ischemic zone was visualized on day 14 using markers

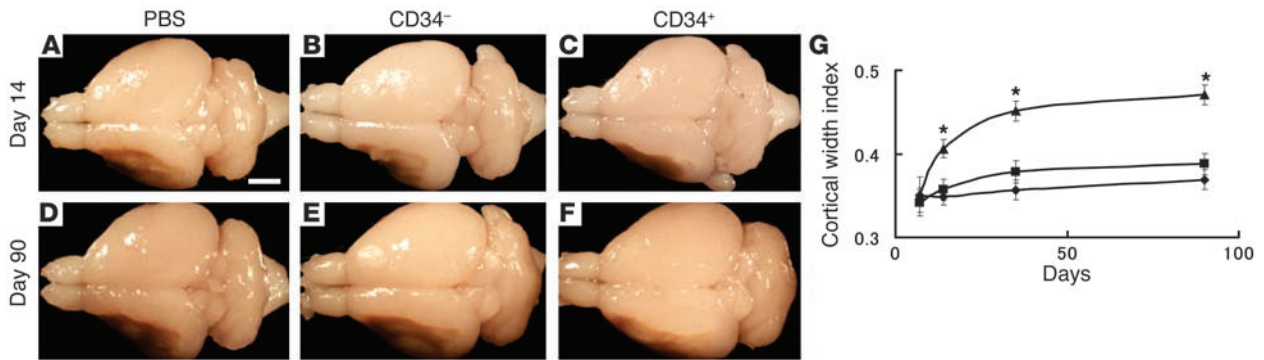


Figure 3

CD34⁺ cell transplantation induces cortical expansion after stroke. (A–F) On day 14 (A–C), and day 90 (D–F) after cell transplantation, the brains of mice were evaluated grossly. Compared with poststroke mice treated with PBS (A and D) or infused with CD34⁻ cells (B and E), animals transplanted with CD34⁺ cells (C and F) showed an increase in area occupied by the left cortex. (G) Cortical regeneration was induced by CD34⁺ cells transplantation: triangles, CD34⁺ cells; squares, CD34⁻ cells; diamonds, PBS. In each case, there were 6 animals per group. Scale bar: 2 mm (A). **P* < 0.05 versus PBS.

for PSA-NCAM (Figure 4, A and B, corresponding to lower and higher magnifications, respectively), as well as the neuronal stem cell marker, Musashi-1 (Figure 4C) (19). Expression of doublecortin (DCX), a marker of immature, or early, neurons (20), was principally restricted to NPCs close to the SVZ (Figure 4D). In contrast, expression of a neuronal cell marker, NeuN, was observed after cell migration from the SVZ and appeared as a signal of lower intensity compared with nearby mature neurons (Figure 4E). The situation was quite different on the contralateral side in poststroke animals treated with CD34⁺ cells; no such movement of precursors (PSA-NCAM⁺ cells) was observed (Figure 4F). Although migrating NPCs were also seen in poststroke animals treated with PBS (Figure 4G) and CD34⁻ cells, this finding was considerably magnified in mice receiving CD34⁺ cells (Figure 4H).

To investigate mechanisms underlying cortical expansion, immunohistological analysis was conducted to visualize PSA-NCAM, NeuN, and MAP-2. On day 14 after transplantation of CD34⁻ cells after stroke, mature cortical neurons displaying neuronal markers NeuN (Figure 5A) and MAP-2 (Figure 5B) were observed up to the periphery of the ischemic area, whereas only a thin layer of migrating PSA-NCAM⁺ NPCs was observed at the ischemic edge (Figure 5C). In contrast, transplantation of CD34⁺ cells after stroke expanded cortical areas displaying a low density of NeuN⁺ (Figure 5D) and MAP-2⁺ cells (Figure 5E) beyond the boundary demarcating mature neurons. Migration of NPCs into this expanded area was also observed by PSA-NCAM staining (Figure 5F).

Next, we investigated the fate of NPCs migrating from the SVZ to the ischemic area. Poststroke animals treated with CD34⁻ cells displayed many cells with TUNEL⁺ nuclei around the lower part of cortical ischemic edge (Figure 5G). In contrast, there were a minimal number of TUNEL⁺ nuclei observed in the same region in animals subject to transplantation of CD34⁺ cells after stroke (Figure

5, H and I). Thus, animals treated with CD34⁺ cells after stroke display migration and survival of NPCs, which eventually contribute to a population of more mature neurons.

To confirm this evidence of neurogenesis, animals transplanted with CD34⁺ cells, CD34⁻ cells, or PBS after stroke were infused with

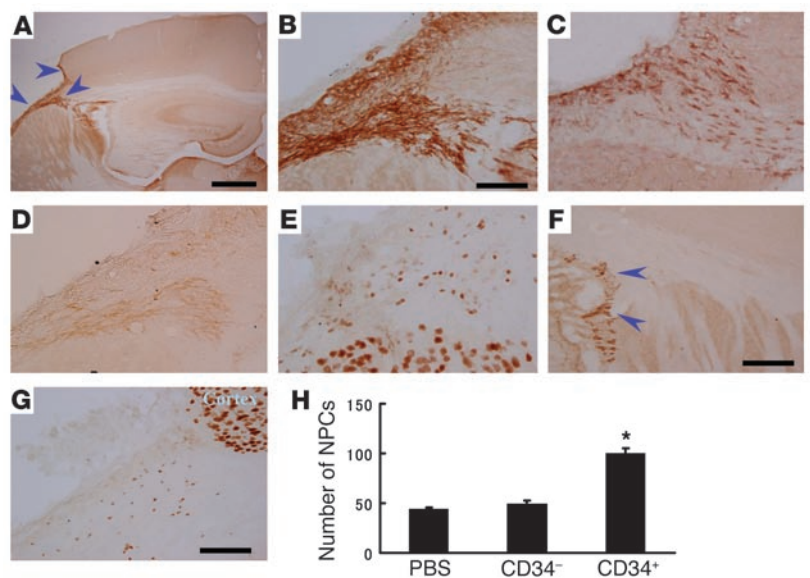
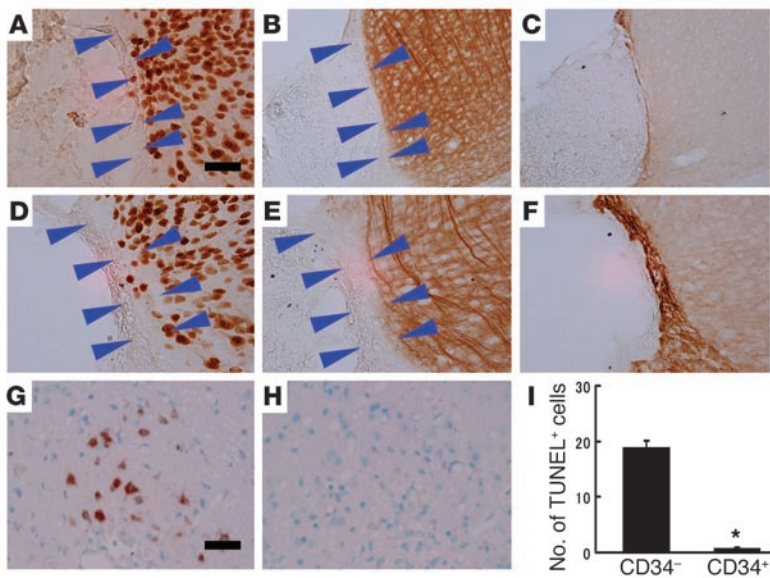


Figure 4

CD34⁺ cell transplantation accelerates neuronal regeneration after stroke. (A and B) On day 14, animals receiving CD34⁺ cells after stroke displayed migration of NPCs toward the ischemic area by PSA-NCAM immunostaining. (C–E) Analysis of serial sections displayed expression of neuronal stem cell markers, Musashi-1 (C) and DCX (D). Note that expression of DCX was limited to the area proximal to the SVZ. PSA-NCAM⁺ NPCs also expressed NeuN (E). Small NeuN⁺ nuclei were observed in PSA-NCAM⁺ NPCs, whereas more intensely staining and larger nuclei represent mature neurons. (F) On day 14 on the contralateral side, PSA-NCAM⁺ NPCs were limited to the SVZ; that is, no migration of NPCs was observed. (G) Migration of NPCs with small NeuN⁺ nuclei toward cortex was observed in poststroke mice treated with PBS on day 14 after cell transplantation. (H) The average number of NPCs in the white matter at the lower left of the left cortex per HPF from 5 animals under each condition. Three sections were evaluated in each animal, and *n* = 5 per group. Arrowheads delineate individual NPCs or demarcate areas rich in NPCs. Scale bars: 1 mm (A), 0.2 mm (B and F), and 0.4 mm (G). **P* < 0.05 versus PBS.

**Figure 5**

Therapeutic neovascularization, due to CD34⁺ cell transplantation after stroke, enhances neurogenesis. (A–F) On day 14 after CD34⁺ cell transplantation, mature cortical neurons were observed up to the edge of the ischemic region displaying neuronal markers, NeuN (A) and MAP-2 (B), whereas only a thin layer of migrating PSA-NCAM⁺ NPCs was observed at the ischemic edge (C). In contrast, after transplantation of CD34⁺ cells, expanded cortical areas displaying a low density of NeuN⁺ (D) and MAP-2⁺ cells (E) were observed beyond the boundary demarcating mature neurons. Migration of NPCs into this expanded area was also observed by PSA-NCAM staining (F). (G–I) On day 14, TUNEL⁺ cells were visualized around the lower part of the expanded cortical area. Whereas massive cell death was observed in animals receiving CD34⁺ cell transplantation (G), the number of TUNEL⁺ profiles was strongly reduced in mice transplanted with CD34⁺ cells (H). (I) The average number of TUNEL⁺ cells per HPF. Three sections were evaluated in each animal; $n = 5$ per group. Arrowheads indicate the expanded cortical areas displaying a low density of indicated marker. Scale bars: 100 μm (A) and 50 μm (G). * $P < 0.05$ for CD34⁺ versus CD34⁺ cell transplantation.

BrdU every second day for a total of 14 days and then killed. BrdU- and NeuN-bearing cell populations were visualized to determine if a group of neurons might be undergoing cell division. In animals treated with PBS alone (Figure 6A) or CD34⁺ cells (Figure 6B), double staining for BrdU and NeuN displayed little evidence of cells (i.e., nuclei) expressing both markers. Only in animals infused with CD34⁺ cells after stroke did we consistently observe double-labeled cell profiles with BrdU and NeuN to produce a merged image with cells displaying both markers (Figure 6C). Quantification of these results showed a significant increase in double-positive profiles (i.e., cells co-staining with antibody to BrdU and NeuN) in poststroke animals infused with CD34⁺ cells (Figure 6D). Consistent with these data, total neuronal counts (Figure 6E) from sections stained with NeuN in the left cortex 90 days after cell transplantation demonstrated a significant increase in the number of cells visualized in animals transplanted with CD34⁺ cells compared with those receiving CD34⁺ cells or PBS. In addition, vasculature was visualized in the ischemic zone of the forebrain of animals treated with CD34⁺ cells 90 days later using anti-mouse CD31 as a marker. A distinct vascular network was seen in the expanded cortex associated with neurogenesis after stroke (Figure 6F). Such vessels displayed a pattern quite distinct from that observed on the contralateral (nonischemic) side (Figure 6G). In these vessels on the ischemic side, some endothelial cells were

observed to express human CD31 antigen (Figure 6H). On day 90, mature MAP-2⁺ dendrites were observed in the expanded cortex of poststroke animals treated with CD34⁺ cells (data not shown). These data are consistent with the formation of a stable neovasculature to support regeneration of cortical neurons.

Inhibition and acceleration of angiogenesis by antiangiogenic and angiogenic agents. To investigate the hypothesis that neovascularization is essential to support endogenous neuronal regeneration, we used an antiangiogenic agent, Endostatin. The latter is known to inhibit proliferation of endothelial cells (21) and to have a direct inhibitory effect on EPCs (22). Mice were subjected to stroke, CD34⁺ or CD34⁺ cells were administered, and animals were divided into two groups; one group received Endostatin and the other PBS ($n = 5$, for each group). On day 7 after cell transplantation, treatment with Endostatin suppressed endothelial proliferation to $18\% \pm 3\%$ and $22\% \pm 3\%$ in mice transplanted with CD34⁺ cells and CD34⁺ cells, respectively, compared with proliferation observed in PBS-treated controls. Administration of Endostatin also impaired cortical expansion due to transplantation of CD34⁺ cells. On day 14, the cortical width index was 0.33 ± 0.01 and 0.41 ± 0.01 with and without Endostatin, respectively ($P < 0.01$). Animals treated with CD34⁺ cells and Endostatin after stroke displayed a reduction in migrating NPCs; 101 ± 7 cells per high-power field (HPF), for animals treated with CD34⁺ cells + PBS, and 21 ± 2 cells/HPF for animals treated with CD34⁺ cells + Endostatin ($P < 0.01$). Similarly, on day 14, the number of neurons in the post-stroke cortex was less in mice treated with CD34⁺ cells + Endostatin ($1,469 \pm 53$), compared with animals treated with CD34⁺ cells + PBS ($2,213 \pm 36$; $P < 0.01$).

To gain further support for our hypothesis, we used the proangiogenic agent erythropoietin (EPO). EPO, well known for its essential role in regulat-

ing proliferation and differentiation of erythroid cells, has recently been found to promote mobilization of EPCs (23) and to have angiogenic potential (24). On day 4 after stroke, a significant increase in circulating CD34⁺ cells was observed with EPO injection ($5,244 \pm 1,267$ and $1,333 \pm 389$ cells/ml in the EPO and PBS groups, respectively; $n = 6$ per group, $P < 0.05$). In addition, mice treated with EPO displayed enhanced neovascularization based on angiographic score (5.4 ± 0.8 and 1.2 ± 0.4 , in EPO and PBS groups, respectively; $n = 5$ per group, $P < 0.05$). On day 28 after stroke, treatment with EPO increased the number of cells co-staining for BrdU and NeuN (3.4 ± 0.3 and 0.4 ± 0.1 /HPF in the EPO and PBS groups, respectively; $n = 5$ per group, $P < 0.05$). It should be noted that the magnitude of the stimulatory effect of EPO treatment on angiogenesis and neurogenesis in these experiments was less than that observed in studies with CD34⁺ cell transplantation, probably because of the greater number of cells transferred in the latter case.

Neurogenesis in 24-week-old mice. To analyze the effect of increased age on accelerated neurogenesis associated with CD34⁺ cell transplantation, 24-week-old SCID mice were used. Compared with poststroke mice treated with CD34⁺ cells (Figure 7A), transplantation of CD34⁺ cells (Figure 7B) showed a significant increase in area occupied by the left cortex on day 35 (Figure 7C). In addi-

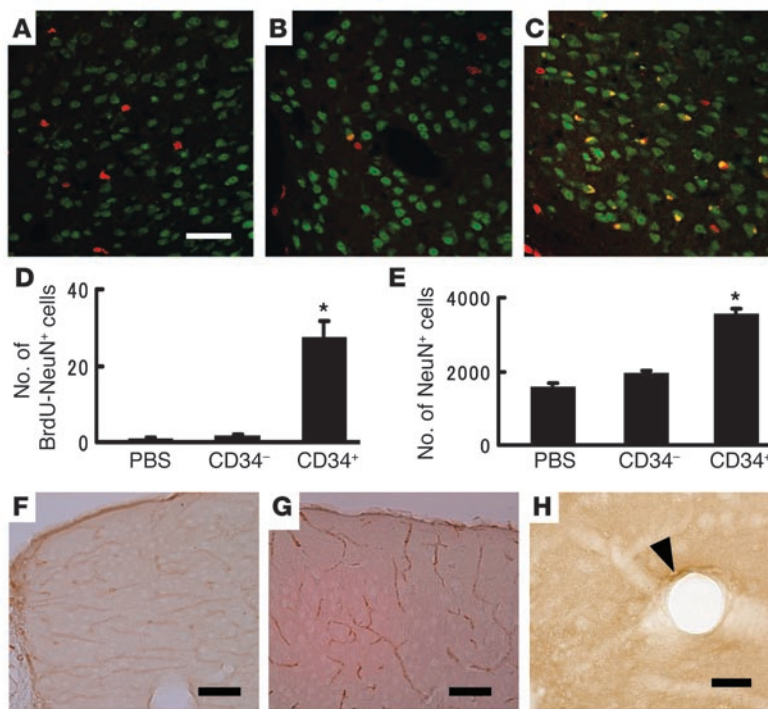


Figure 6 Therapeutic neovascularization supports survival of regenerating neurons. (A–C) Animals treated with PBS alone (A), CD34⁻ cells (B), or CD34⁺ cells (C) were infused with BrdU, killed, and studied immunohistochemically with antibody to BrdU (red), NeuN (green), or both (yellow). (D) The average number of double-positive cells (stained with antibody to BrdU and NeuN) per HPF on day 90 after cell transplantation. Ten fields were evaluated in each animal, and *n* = 6 per group. **P* < 0.05 versus PBS. (E) On day 90 after cell transplantation, brain sections from animals treated with PBS alone, CD34⁻ cells, or CD34⁺ cells were stained with antibody to NeuN, and the number of total neurons in the left cortex was counted. (E) The average number of total NeuN⁺ cells in the left cortex. *n* = 6 per group; **P* < 0.05 versus PBS. (F and G) On day 90 after cell transplantation, mouse CD31 was visualized immunohistologically in forebrain sections from poststroke animals. Newly formed vascular networks were observed in the expanded cortex. The vascular pattern displayed by these neovessels on the ipsilateral (F, ischemic side) was different from that observed on the contralateral side (G). (H) On day 90, human CD31 was visualized immunohistologically. Human endothelial cells were observed in the regenerating cortex of animals treated with CD34⁺ cells after stroke. The arrowhead shows a human CD31⁺ endothelial cell. Scale bars: 50 μm (A), 100 μm (F and G), and 20 μm (H).

tion, evidence of vascular activation, as judged by reactivity with anti-CD13 antibody, was observed in mice treated with CD34⁺ cells (Figure 7D, CD34⁻ cells; Figure 7E, CD34⁺ cells). Migration of NPCs was observed in mice treated with CD34⁻ cells (Figure 7F) and CD34⁺ cells (Figure 7G), but a significant enhancement was observed in animals receiving CD34⁺ cells (Figure 7H). Only a thin layer of migrating PSA-NCAM⁺ NPCs was observed at the ischemic edge in animals treated with CD34⁻ cells (Figure 7I). In contrast, CD34⁺ cell transplantation resulted in a much thicker layer of PSA-NCAM⁺ cells (Figure 7J). Comparing these data in 24-week-old animals with our previous results using 5-week-old mice, the

older animals displayed a similar cortical width index (5 weeks old, 0.45 ± 0.01; 24 weeks old, 0.43 ± 0.02; *P* > 0.05), although older age was associated with a decrease in NPCs migrating into the cortex (5 weeks old, 71 ± 3 cells/HPF; 24 weeks old, 43 ± 4 cells/HPF; *P* < 0.05) on day 35 after CD34⁺ cell transplantation.

Discussion

Our findings show that administration of human CD34⁺ cells to immunocompromised mice 48 hours after stroke enhances neovascularization at the border of the ischemic zone followed by endogenous neurogenesis. Furthermore, suppression of the neo-

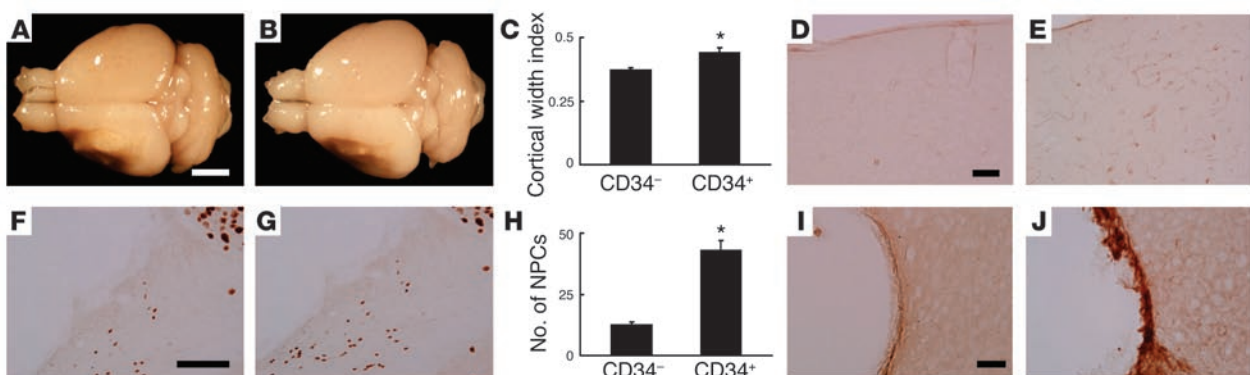


Figure 7 CD34⁺ cell transplantation in 24-week-old animals. (A–C) On day 35 after cell transplantation, the brains were evaluated. Compared with post-stroke mice treated with CD34⁻ cells (A), animals transplanted with CD34⁺ cells (B) showed an increase in area occupied by the left cortex. Significant cortical regeneration was induced by CD34⁺ cells transplantation (C). (D and E) Compared with CD34⁻ cell transplantation (D), increased evidence of activated vasculature was observed in animals receiving CD34⁺ cells (E), as detected with mouse specific anti-CD13 antibody. (F–H) Migration of NPCs (small NeuN⁺ nuclei migrating toward the cortex) was observed in poststroke mice treated with CD34⁻ cells (F) and with CD34⁺ cells (G). However, a significant increase in migrating NPCs was induced by CD34⁺ cell transplantation (H). (I and J) A thin layer of migrating PSA-NCAM⁺ NPCs was observed at the ischemic edge of the cortex in animals treated with CD34⁻ cells (I), compared with a much thicker layer in those receiving CD34⁺ cells (J). Scale bars: 2 mm (A), 0.1 mm (D and I), and 0.4 mm (F). *n* = 4 in each group; **P* < 0.05 versus CD34⁻ cells.



vascularization by an antiangiogenic agent impaired neurogenesis. On the basis of these data, accelerated neovessel formation seems to be essential for enhancing endogenous neurogenesis and improving functional recovery.

The generation of new neurons in the adult is largely restricted to two regions: the SVZ lining the lateral ventricles and the subgranular zone of the dentate gyrus (25). In the transient cerebral ischemia model, evidence has been provided that neuronal regeneration occurs (10, 20). However, even in a transient ischemia model in which integrity of the microcirculation is maintained, it was shown that greater than 80% of newly formed neurons died, most likely because of unfavorable environmental conditions including lack of trophic support and exposure to products of damaged tissue. These considerations may underlie the observation that only 0.2% of nonviable ischemic neurons were replaced through neurogenesis (20). Such previous observations are consistent with our current results in animals treated with CD34⁺ cells or PBS, in which there was no enhancement of neovascularization, no neurogenesis, and no functional recovery.

Consistent with these data, Endostatin-mediated suppression of endothelial proliferation and direct effects on EPCs abrogated the beneficial effect CD34⁺ cells on neurogenesis and cortical expansion, and at the same time inhibited formation of neovasculature after stroke. In contrast, when EPO was used as a proangiogenic agent (recently, EPO has also been found to promote mobilization of EPCs) (23, 24), accelerated formation of neovasculature was accompanied by enhanced neurogenesis after stroke in our murine model. Our data provide the first direct link between therapeutic neovascularization after stroke and enhanced neurogenesis; formation of neovasculature after stroke supported neurogenesis. Consistent with a previous report (26), activation of NPCs after stroke was induced in adult, as well as young, murine brain. The number of activated NPCs was less in the adult animals, but a significant increase was still observed consequent to CD34⁺ cells transplantation, with evidence of activated vasculature in brains of the older animals. These findings support our hypothesis that administration of CD34⁺ cells provides a milieu favorable for neovascularization and endogenous neurogenesis, even in the mature brain.

A relationship between angiogenesis and neurogenesis would be consistent with regeneration of parenchymal cells in other organs subject to therapeutic angiogenesis (27, 28). Mechanisms underlying this observation might include more optimal preparation of the ischemic tissue bed for neuronal regeneration by accelerated removal of debris and toxic products, and/or the enhanced production of chemokines and trophic agents by neovasculature. Factors involved might be FGF2 (11), PDGF (29), brain-derived neurotrophic factor (30), and IL-8 (31), which have the capacity to induce mitogenesis, differentiation, recruitment, and survival of NPCs and newly generated neurons. In addition, mediators produced by CD34⁺ cells (4), such as VEGF, FGF2, and IGF-1, have been shown to accelerate endogenous neurogenesis (10–12). The effects of these and other factors derived from CD34⁺ cells (32, 33) acting on the vasculature are also likely to have an important role in providing an environment conducive to neurogenesis. Newly formed neurons in the setting of neovascularization have been shown to integrate into neuronal networks in adult animals (34); in fact, lack of participation of neurons in such neuronal circuits is probably associated with cell death (20). Our results provide strong support for the hypothesis that neovascularization consequent to administration of CD34⁺ cells induces neurogenesis by providing the necessary supportive environment. Whereas perturbation of the neurovascular unit has been proposed to contribute to tissue dam-

age in stroke, neoangiogenesis and accompanying neurogenesis could be considered to be rebuilding crucial elements of the neurovascular unit (35). Previous reports have demonstrated that endothelial cells in newly formed vessels after stroke mainly originate from pre-existing endothelial cells (due to proliferation) with a contribution of circulating EPCs (36, 37). Consistent with these findings, we observed proliferating mouse endothelial cells around the ischemic area soon after stroke. The latter endothelial cells could be visualized, at least in part, with antibody specific for human CD31, after transplantation of human CD34⁺ cells. Furthermore, diminished neovascularization was observed after stroke following transplantation of CD34⁺/Flk-1⁻ cells, compared with CD34⁺ cells (containing both Flk-1⁻ and Flk-1⁺ cells) despite similar activation of murine endothelial cells in the ischemic territory in each case. These results support potential contribution of CD34⁺/Flk-1⁺ cells (a population known to be rich in EPCs) in expansion of the vascular network after stroke.

The actual pattern of the newly formed vascular network was quite different on the ipsilateral (ischemic) from the contralateral side in poststroke mice treated with CD34⁺ cells. This observation suggests that formation of vasculature consequent to transplantation of CD34⁺ cells after stroke does not simply reconstitute the original vascular network. Rather, a new vascular pattern arises (a true neovasculature) that is capable of supporting neurogenesis, followed by functional recovery, even though it displays a relatively “aberrant” pattern, at least anatomically, compared with the contralateral vasculature.

Our observations provide evidence of a crucial role for neovessel formation, achieved through the administration of CD34⁺ cells after stroke, in processes that underlie neurogenesis. These data strongly suggest that neovascularization is essential for neuronal regeneration after stroke and that therapeutic neovascularization is a potentially effective means of enhancing functional recovery. Our observations might explain the mild therapeutic effect achieved by neuronal cell transplantation after stroke reported in humans (38). This leads to the hypothesis that therapeutic neovascularization may be required to achieve optimal “take” of transplanted neuronal precursors in the setting of ischemia. Although the current data bear most directly on the endogenous neuronal response to cerebrovascular ischemia, it is possible that enhanced formation of neovasculature may also be important for survival of embryonic (39) and neural (40) stem cell transplants in other circumstances, such as neurodegenerative disorders.

Methods

All procedures were done in accordance with the National Cardiovascular Center Animal Care and Use, and Human Assurance Committees. Quantitative measurements and behavioral tests were conducted by investigators blinded to the experimental protocol and identity of the sections and animals under study.

Induction of focal cerebral ischemia. Permanent focal cerebral infarction was produced by ligation and disconnection of the distal portion of the left MCA. Male SCID mice (5 weeks old; Oriental Yeast Co. Ltd., Tokyo, Japan) were used for experiments involving human cell transplantation. Male C57BL/6J mice (24 weeks old; Clea Japan Inc., Tokyo, Japan) were used for experiments with human recombinant EPO (Kirin, Tokyo, Japan). Under halothane inhalation (3%), the left MCA was isolated, electrocauterized, and disconnected just distal to its crossing of the olfactory tract (distal M1 portion). CBF in the MCA area was monitored as described (41). Mice that showed decreased CBF by approximately 75% immediately after and 24 hours after ligation were used for our experiments (success rate of >95%). Body temperature was maintained at 36.5–37°C using a heat lamp during



the operation and for 2 hours after MCA occlusion. The ratio of CBF after cell transplantation to CBF before cell transplantation was calculated to evaluate the change of CBF (termed relative CBF).

Analysis of perfusion and brain infarction after MCA occlusion. Carbon black ink (Fuekinori Kogyo Co. Ltd., Osaka, Japan) was used to delineate the perfused area after MCA ligation as described (42). Viability of brain tissue was evaluated using TTC (Sigma-Aldrich, St. Louis, Missouri, USA). Coronal sections (1 mm thick) of the forebrain were stained with 1% TTC, and the images of sections from the exact center of the forebrain were captured using a microscopic digital camera system (Olympus, Tokyo, Japan). The width of viable cortex was determined by NIH image.

Administration of CD34⁺ cells after stroke. CD34⁺ (containing Flk-1⁻ and Flk-1⁺ cell populations) and CD34⁺/Flk-1⁻ cells were isolated from cord blood using a Direct CD34 Progenitor Cell Isolation kit (Miltenyi Biotec, Bergisch Gladbach, Germany), CD34 Multisort kit (Miltenyi Biotec), and anti-Flk-1 antibody (Sigma-Aldrich), according to the manufacturers' protocols. CD34⁺ mononuclear cells were also collected for control experiments. In each case, cell populations were analyzed by FACS (BD Biosciences, San Jose, California, USA) using PerCP-conjugated anti-CD34 antibody (BD Biosciences), phycoerythrin-conjugated (PE-conjugated) anti-PIH12 antibody (BD Biosciences), and anti-Flk-1 antibody (Sigma-Aldrich) conjugated with FITC according to the manufacturer's protocol (Zenon Alexa Fluor; Molecular Probes Inc., Eugene, Oregon, USA). For labeling studies, cells were incubated with the fluorescent dye chloromethylbenzamide (CM-DiI; Molecular Probes Inc.). Forty-eight hours after stroke, 5×10^5 CD34⁺, CD34⁺/FLK-1⁻, CD34⁻ cells, or the same volume (100 μ l) of PBS were infused intravenously via the tail vein.

For the analysis of the effect of aging on neurogenesis, 24-week-old male SCID mice were used. Because of the known increased evidence of tumors, infection (for example, excess mucus in the eye), and elevated plasma immunoglobulins in older SCID mice, the presence of each of these conditions was also investigated. Mice with evidence of weight loss, tumors, infection, or elevated plasma immunoglobulins (>10 μ g/ml) were excluded from further analysis.

Immunohistochemistry. Mice brains were removed and fixed in paraformaldehyde. Coronal sections (20 μ m) were prepared using a vibratome (Leica Microsystems Inc., Wetzlar, Germany). To assess cell proliferation and angiogenesis, fresh frozen sections (20 μ m) were prepared and fixed in acetone. Sections were subjected to immunohistochemistry with antibody to NeuN (Chemicon International, Temecula, California, USA), mouse-specific antibody to CD31 (BD Biosciences), human-specific antibody to CD31 (DAKO A/S, Glostrup, Denmark), mouse-specific antibody to CD13 (Santa Cruz Biotechnology, Santa Cruz, California, USA), and antibodies to MAP-2 (Sigma-Aldrich), human von Willebrand Factor (DAKO A/S), PSA-NCAM (Chemicon International), DCX (Chemicon International), Musashi-1 (19), TUNEL (Molecular Probes Inc.), or BrdU (Boehringer Ingelheim, Ingelheim, Germany).

Cell proliferation analysis. For assessment of cell proliferation *in vivo*, BrdU (Sigma-Aldrich) was administered intraperitoneally. In the case of endothelial cell proliferation, BrdU (200 mg/kg) was administered 24 hours before killing. For endogenous neurogenesis, 50 mg/kg of BrdU was injected every second day for a total of 14 days. Cells co-staining for BrdU and mouse-specific antibody to CD31 using confocal microscopy (Olympus) in HPFs ($\times 40$ of objective lens) were counted as proliferating endothelial cells. The cell population co-staining for BrdU and NeuN was counted as regenerating-proliferating neurons. For quantitative analysis, brain sections at the exact center of the forebrain were stained, and the left cortical area of 1 mm (or >1 mm) distal from the midline was examined by two investigators blinded to the experimental protocol. In each case, 10 representative fields were evaluated before quantitative results were analyzed.

Assessment of angiogenesis. For assessment of angiogenesis at the border of MCA and ACA areas, carbon black perfusion was used to visualize the

vasculature and TTC staining to demarcate the border of viable and non-viable tissue. Semi-quantitative analysis of angiogenesis made use of the angiographic score and was calculated by a previously described method (43). Microscopic digital images were scanned into a computer (Olympus), and an overlay, composed of 50- μ m-diameter circles arranged in rows spaced 100 μ m apart, was superimposed on the image. The total number of grids intersecting the border zone containing a microvessel was counted. Mouse-specific antibody to CD13 was used to visualize 'activated' vasculature.

Assessment of ischemic brain damage and neurogenesis. For determination of the cortical width index, whole-brain images were captured using a microscopic digital camera system (Olympus). The width at the midpoint of the forebrain was measured, and the ratio of left width to right width was defined as the cortical width index. To quantify accelerated neurogenesis, three brain sections around the center of the forebrain on day 14 after cell transplantation were stained with NeuN or TUNEL. The number of small NeuN nuclei-positive cells in the white matter at lower left of the left cortex and the number of TUNEL⁺ cells around the lower part of ischemic cortical edge was counted in HPFs. For determining the number of neurons in the left cortex, brain sections of the exact center of the forebrain on day 90 were stained with NeuN, and the total number of NeuN⁺ cells was quantitated.

Behavioral analysis. To assess cortical function, mice were subjected to behavioral testing in the open-field task (44) on day 35 or 90 after cell transplantation. Auditory startling behavior was tested as described (45). To estimate learning activity, mice were tested by water maze test (46) and passive avoidance test (47). To exclude the contribution of physical deficits directly related to the operative procedure and induction of stroke, motor deficiencies were examined on days 9 and 16 after stroke. Neurological deficits were scored on a three-point modified scale as described (17). Body weight, monitored in each experimental group, displayed no significant differences (data not shown).

Inhibition and promotion of neovascularization. Inhibition of neovascularization was achieved by administering Endostatin (10 μ g; subcutaneous; Calbiochem-Novabiochem, Darmstadt, Germany) once daily for 14 days. To stimulate mobilization of EPCs originating in the host, EPO (1,000 μ g/kg) was injected subcutaneously 24, 48, and 72 hours after ischemia. Peripheral blood cells were analyzed using FITC-conjugated anti-mouse CD45 (BD Biosciences), PE-conjugated anti-mouse CD34 antibodies (BD Biosciences), and 7-amino-actinomycin D viability Dye (Beckman Coulter Inc., Marseille Cedex, France), as described (48), using counting beads (BD Biosciences) as an internal control.

Data analysis. In all experiments, mean \pm SE is reported. Statistical comparisons among groups were determined using one-way ANOVA. Where indicated, individual comparisons were performed using Student's *t* test.

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